

# High-resolution crystal structure of LINE-1 Reverse Transcriptase and Its Implications for Rational Inhibitor Design

**ROME**  
THERAPEUTICS



View this poster on our website

**Authors:** Eric T. Baldwin<sup>1</sup>, Trevor van Eeuwen<sup>2</sup>, David Hoyos<sup>3</sup>, Arthur Zalevsky<sup>4</sup>, Egor P. Tchesnokov<sup>5</sup>, **Roberto Sanchez**<sup>1</sup>, Luciano DiStefano<sup>6</sup>, Francesc Xavier Ruiz<sup>7</sup>, Matthew Hancock<sup>4</sup>, Thomas Walpole<sup>8</sup>, Charles Nichols<sup>8</sup>, Paul Wan<sup>8</sup>, Kirsi Riento<sup>8</sup>, Rowan-Halls Kass<sup>8</sup>, Martin Augustin<sup>9</sup>, Alfred Lammens<sup>9</sup>, Anja Jester<sup>9</sup>, Paula Upla<sup>2</sup>, Kera Xibinaku<sup>10</sup>, Samantha Congreve<sup>10</sup>, Maximiliaan Hennink<sup>10</sup>, Kacper B. Rogala<sup>10, 11</sup>, Anna M. Schneider<sup>12</sup>, Jennifer E. Fairman<sup>13</sup>, Shawn M. Christensen<sup>14</sup>, Wenyan Miao<sup>1</sup>, Dennis M. Zaller<sup>1</sup>, Andrej Šali<sup>4</sup>, Oliver Weichenrieder<sup>12</sup>, Kathleen H. Burns<sup>15</sup>, Matthias Götte<sup>5</sup>, Michael P. Rout<sup>2</sup>, Eddy Arnold<sup>2</sup>, Benjamin D. Greenbaum<sup>3, 16</sup>, Donna L. Romero<sup>1</sup>, John LaCava<sup>2, 6</sup> and Martin S. Taylor<sup>17</sup>

**Affiliations:** 1 - ROME Therapeutics, Boston MA, USA.; 2 - Laboratory of Cellular and Structural Biology, The Rockefeller University, New York, NY, USA.; 3 - Computational Oncology, Department of Epidemiology & Biostatistics, Memorial Sloan Kettering Cancer Center, New York, NY, USA.; 4 - Department of Bioengineering and Therapeutic Sciences and Department of Pharmaceutical Chemistry, Quantitative Biology Institute, University of California, San Francisco, San Francisco, CA, USA.; 5 - Department of Medical Microbiology and Immunology, University of Alberta, Edmonton, Alberta, Canada.; 6 - European Research Institute for the Biology of Ageing, University Medical Center Groningen, Groningen AV, The Netherlands.; 7 - Center for Advanced Biotechnology and Medicine and Department of Chemistry and Chemical Biology, Rutgers University, Piscataway, NJ USA.; 8 - Charles River Laboratory, Chesterford Research Park, United Kingdom.; 9 - Proteros Biostructures GmbH, Planegg-Martinsried, Germany.; 10 - Whitehead Institute for Biomedical Research, Cambridge, MA, USA.; 11 - Stanford Cancer Institute, Department of Structural Biology, and Department of Chemical and Systems Biology, Stanford University, Palo Alto, CA, USA.; 12 - Structural Biology of Selfish RNA, Department of Protein Evolution, Max Planck Institute for Developmental Biology, Tübingen, Germany.; 13 - Johns Hopkins University School of Medicine, Baltimore, MD, USA.; 14 - Department of Biology, University of Texas at Arlington, Arlington, TX, USA.; 15 - Department of Pathology, Dana Farber Cancer Institute and Harvard Medical School, Boston, MA, USA.; 16 - Physiology, Biophysics & Systems Biology, Weill Cornell Medicine, Weill Cornell Medical College, New York, NY, US.; 17 - Department of Pathology, Massachusetts General Hospital, and Harvard Medical School, Boston, MA USA

## Background

The LINE-1 (L1) retrotransposon has written almost half of the human genome through a “copy-and-paste” mechanism catalyzed by its multifunctional enzyme ORF2p, which contains a reverse transcriptase (RT) domain at its core. LINE-1 RT has emerged as a potential therapeutic target implicated in cancer, autoimmunity, and aging. However, a lack of structural and mechanistic knowledge hampers efforts to rationally exploit it. Here, we report the first high-resolution crystal structure of the human LINE-1 RT bound to a template-primer duplex and incoming deoxynucleotide triphosphate (dNTP) and characterize its inhibition profile by existing RT inhibitors.

## LINE-1 Reverse Transcriptase (RT) structure and its relationship to other RT domains

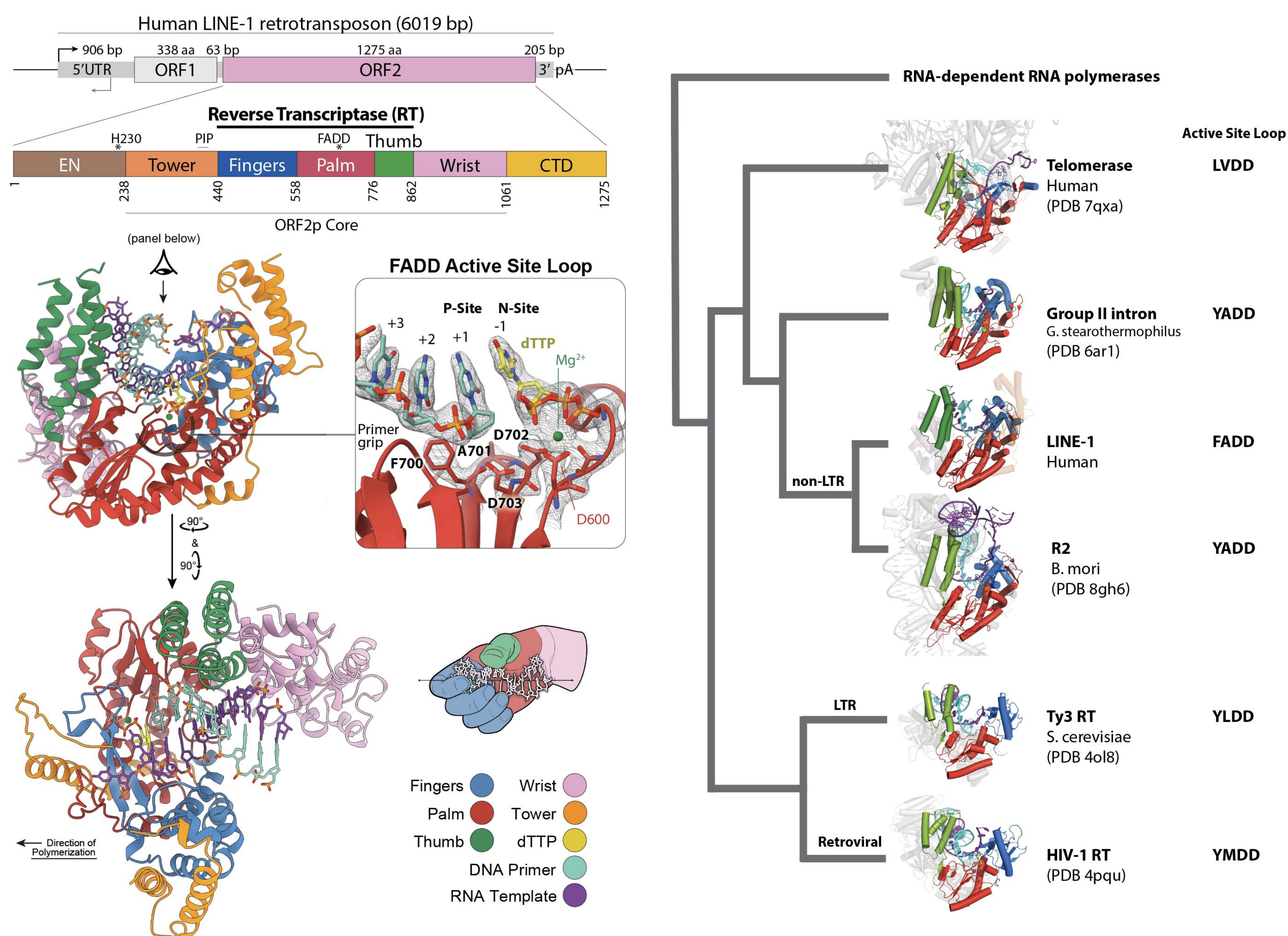


Figure 1. (Left) Structure of human LINE-1 ORF2p core containing the Reverse Transcriptase domain. (Right) Comparison of evolutionarily related RT domains of known structure (tree adapted from Xiong & Eickbush, 1990).

## LINE-1 RT is not inhibited by HIV-1 Non-Nucleoside RT inhibitors (NNRTi)

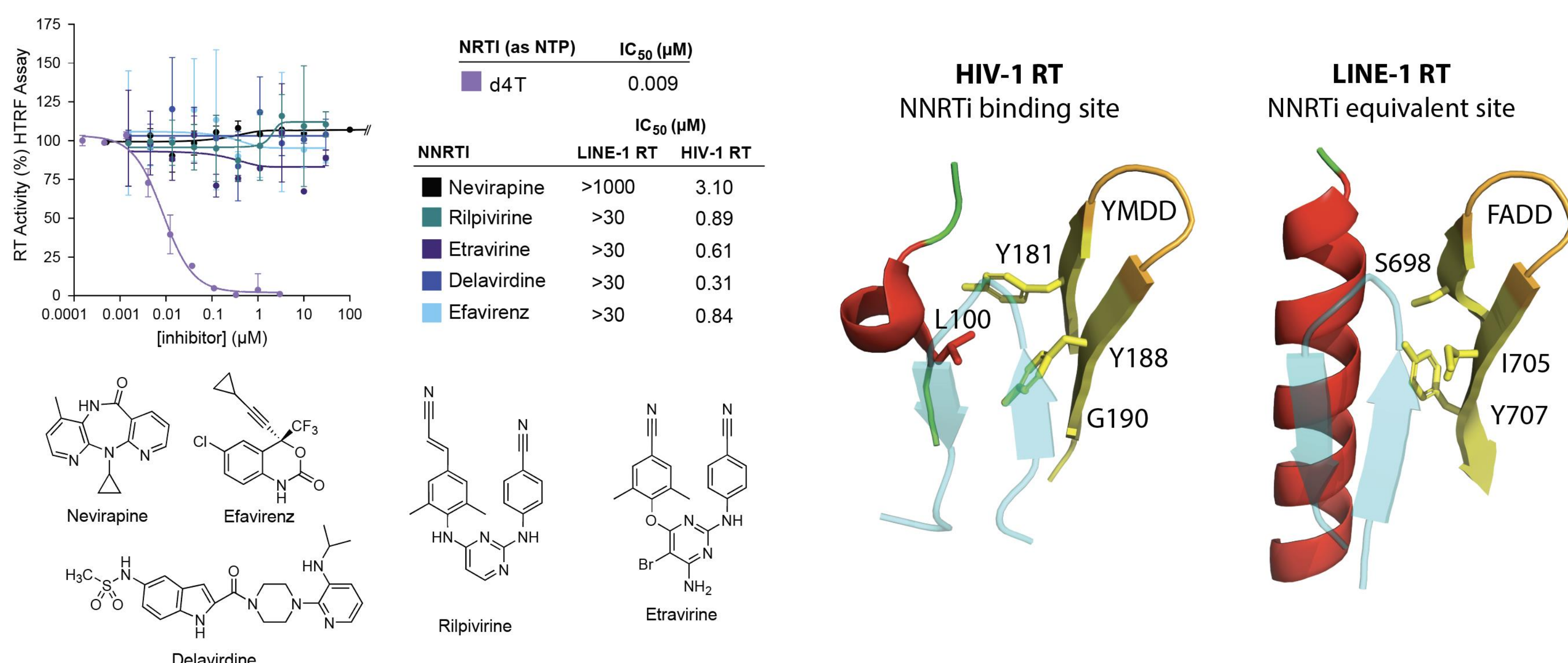


Figure 2. (Left) Inhibition of LINE-1 RT by HIV-1 NNRTi. (Right) Comparison of HIV-1 NNRTi binding site and the equivalent region in LINE-1 RT. The HIV-1 NNRTi site is not conserved in LINE-1 RT.

## LINE-1 RT inhibition by Nucleoside RT inhibitors (NRTi) differs from HIV-1 RT and Telomerase

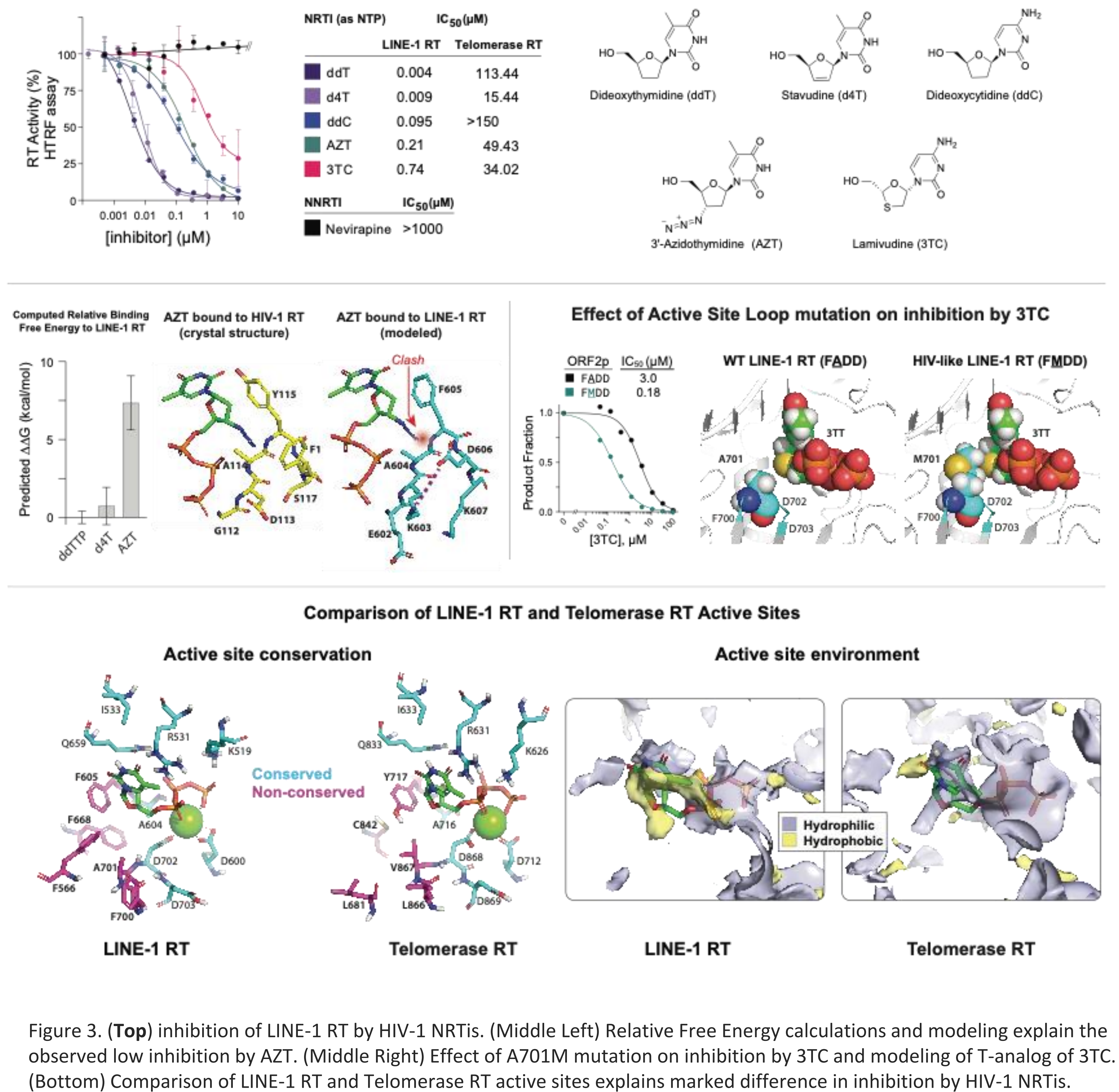


Figure 3. (Top) inhibition of LINE-1 RT by HIV-1 NRTi. (Middle Left) Relative Free Energy calculations and modeling explain the observed low inhibition by AZT. (Middle Right) Effect of A701M mutation on inhibition by 3TC and modeling of T-analog of 3TC. (Bottom) Comparison of LINE-1 RT and Telomerase RT active sites explains marked difference in inhibition by HIV-1 NRTi.

## Results

- The LINE-1 RT crystal structure confirms the closer evolutionary relationship of non-LTR retroelement RTs with bacterial RTs and reveals marked differences with retroviral and LTR retroelement RTs.
- Characterization of LINE-1 RT inhibition by nucleoside and non-nucleoside HIV-1 RT inhibitors reveals a unique inhibition profile for LINE-1 RT, showing lower inhibition by some potent HIV-1 inhibitors such as AZT and 3TC, and no inhibition by non-nucleoside RT inhibitors.
- Computational modeling reveals that the crystal structure can provide qualitative and quantitative explanations for the differences in inhibition between LINE-1 RT and HIV-1 RT, as well as the global differences in inhibition between LINE-1 RT and Telomerase RT.

## Conclusion

The determination of the first high resolution crystal structure of LINE-1 Reverse Transcriptase and testing of HIV-1 inhibitors against it reveals that LINE-1 RT has structural properties that translate into a unique inhibition profile that differs from HIV-1 RT. Development of LINE-1 RT specific inhibitors will require a dedicated effort that is enabled by the newly determined crystal structure.